



EVALUATION OF EXON 17 OF INSULIN RECEPTOR (INSR) GENE AND ITS RELATIONSHIP WITH DIABETES TYPE 2 IN AN IRANIAN POPULATION

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Abolhasan REZAEI^{1*}, Sheyda AKHSHABI¹, FARIBA SADEGHI²

¹Department of Genetics–School of Basic Science, Islamic Azad University, Tonekabon Branch, IRAN

² Department of Medicine, Islamic Azad University, Tonekabon Branch, IRAN

Corresponding Author: Abolhasan Rezaei, e-mail: a.rezaei@tonekaboniau.ac.ir

Abstract. Mutations in insulin receptor gene cause the inherited insulin resistant syndrome, especially diabetic diseases. Here we optimized the conditions for sequencing of a partial fragment of INSR gene, exon 17. We sequenced fourteen fragments from a diabetic sample and a control group chosen from an Iranian population. In this study we analyzed eleven sequences of diabetic's patients by DNAMAN program (DEMO 8.0), deposited in Genbank with accession numbers LC055416, LC055417, LC055419, LC055421, LC055423, LC055424, LC055425, LC055426, LC055496, LC055497, and LC055498. From our control group, three sequences were deposited in Genbank with accession numbers of LC055418, LC055420, and LC055422. Results showed that there were variation between the sequences of exon 17 of INSR gene in diabetics and control group. Variations were observed in fragments at the beginning and the end of exon 17 among diabetics population. Moreover we found subjected SNPs between diabetic's patients that haven't been reported by other researchers. In this study we concluded that mutations in exon 17 of INSR gene contributed in diabetic diseases.

Keyword: Insulin receptor gene, Exon 17, Diabetes, Sequencing, Control group

Introduction

Insulin receptor (INSR) gene is consisted of 22 exons spanning 120 kilobases on chromosome 19 [KADOWAKI *et al.*, 1990; LI *et al.*, 2006; MOELLER *et al.*, 1990].

Studies were showed that mutations in the insulin receptor gene cause the inherited insulin resistant syndromes, Leprechaunism and Rabson–Mendenhall syndrome [LONGO *et al.*, 1994].

These recessive conditions are characterized by intrauterine and post-natal growth restrictions, dysmorphic features, altered glucose homeostasis, and early demise.

The region of exons 17–21 encodes the tyrosine kinase domain of the receptor, which is necessary for insulin signal transduction.

Mutations in exons 17–21 are very important because they are the causes of insulin resistance and hyperinsulinemia [SINO *et al.*, 1990; HOFFMAN *et al.*, 1997; WARD *et al.*, 2007].

Researchers had shown that SNPs in exon 17 to 21 had a relationship with INSR β subunit gene in women with

PCOS. Talbot and collab. [TALBOT *et al.*, 1996] using Southern blot analysis showed that a major mutation in INSR contributed with PCOS.

Other authors [SIEGEL *et al.*, 2002; DIAO *et al.*, 2004, LEE *et al.*, 2006], had reported that polymorphism in INSR induces mild changes in INSR function which may contribute to the development of PCOS.

PCOS in women were observed almost between 5–10 % of them when they reached to the age of puberty.

The insulin receptor itself may be a susceptibility gene for PCOS [TALBOT *et al.*, 1996; TUCCI *et al.*, 2001; WARD *et al.*, 2007; DUNAIF, 2012; KNOCHENHAUER *et al.*, 1998; SAN MILAN *et al.*, 2004].

Insulin receptor gene has twenty two exons and twenty one introns that are located on chromosome 19.

Domains of receptor tyrosine kinase on exons 17 to 21 are encoded to convey the message (signal transduction), which is necessary for function of insulin hormone.



Mutations in exon 17 cause insulin resistance and lead to hyperinsulinemia [XITA *et al.*, 2003; WEEDON *et al.*, 2003; LEE *et al.*, 2006].

In this research we studied the exon 17 of INSR gene in diabetic patients and control groups, and based on variations among them, we tried to find a number of SNPs in diabetic patients that could be the cause of diabetic diseases in an Iranian population.

Material and methods

Sample collection

Sixty samples of diabetic patients and control groups were collected from an Iranian population (age 32.34 ± 3.1 years old with a BMI of 26.23 ± 3.32 for patients and 31.56 ± 2.41 KG/m^2 mean \pm SD for control group, approximately) recruited from endocrinology clinics in north of Iran.

(The BMI was calculated by dividing the weight in kilograms by squared height in meters).

DNA extraction

DNA was extracted with phenol–chloroform method using proteinase K according to the protocol of Sambrook [SAMBROOK *et al.*, 1998; BUTNARIU *et al.*, 2016].

Genomic DNA was isolated in both groups (diabetic patients and control group) and used to determine the functions of exon 17 in INSR gene.

For all groups we designed one pair of primers as follows:

INSR exon 17 Forward

Tgtaaacgacgcccagtcgaaggatgctgtgtagataag

INSR exon 17 Reverse

caggaaacagctatgacctcaggaaagccagcccatgtc

These primers were applied in a PCR program and the reaction for getting a sharp band on the gel electrophoresis was as follows:

PCR amplification for Human insulin receptor gene – exon 17:

Template DNA (100ng/ μL)	3.0 μL
Forward primer (10pmol/ μL)	0.5 μL
Reverse primer (10pmol/ μL)	0.5 μL
dNTP mix (2.5mM each)	1.0 μL
10X buffer	2.5 μL
MgCl ₂	2.5 μL
Taq enzyme (3U/ μL)	0.3 μL
Water	15 μL
Total Reaction volume	25.0 μL

PCR Cycle condition:

94°C	95°C	55.5°C	72°C	72°C
5 min	30 sec	40 sec	30sec	10 min
40 cycles				

PCR products were run on 1.5 percent gel electrophoresis to determine the quantity and quality of PCR products. PCR products were purified by High Pure PCR Product purification kit (cat no. 234.32, Cinagene Company).

Purified samples of suspected cases of diabetes and control samples were sent for sequencing (It should be noted suspicious samples of these patients were selected on the basis of a questionnaire).

Results and discussion

The PCR products were separated by electrophoresis in a TBE agarose gel containing ethidium bromide using standard protocols.

The desired PCR product band was visualized in a medium or long wavelength (e.g., ≥ 300 nm) UV light, and excised quickly to minimize the exposure of DNA to UV light.

The PCR product was transferred to a 1.5 ml micro centrifuge tube for sequencing.

Only samples with good concentration were selected and subjected to sequencing.

In Figure 1, sequences of exon 17 INSR gene in control group with accession numbers LC055418.1, LC055420 and LC055422.1 were analyzed by DNAMAN program.

According to our result, except for the first 24 nucleotides, there was high homology between sequences (100 %).

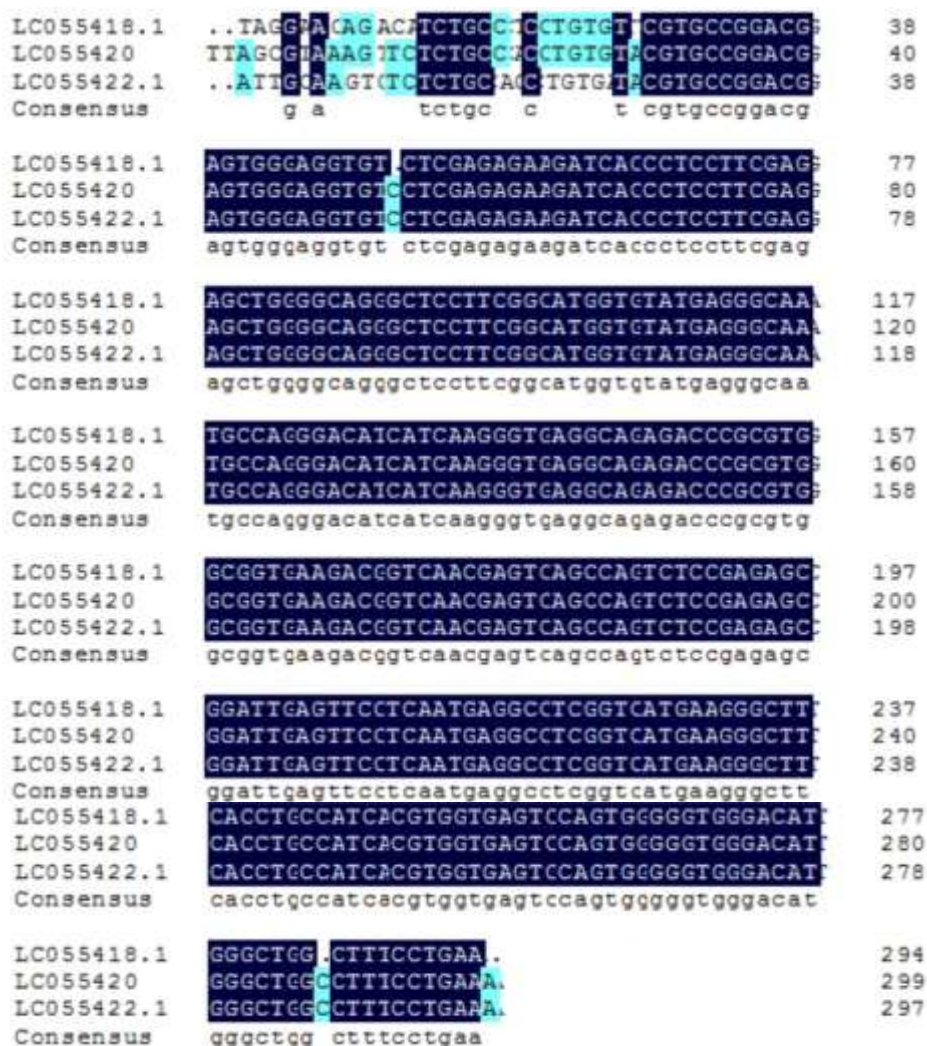


Figure 1: Results between the sequences of exon 17 in INSR gene in control group accession nos. LC055418.1, LC055420 and LC055422.1 by DNAMAN program. According to our results, except for the first 24 nucleotides, there was high homology between sequences (100 %).

In **Figure 2** sequences of exon 17 in INSR gene were compared by DNAMAN program in diabetic patients with accession numbers of:

LC055416,
LC055417,
LC055419,
LC055421,
LC055423,
LC055424,
LC055425,
LC055426,
LC055496,
LC055497,
LC055498.

Based on our results, except for first 21 nucleotides and the last 12 nucleotides at the end of exon 17 INSR gene, other region of exon 17 has high homology between them.

To understand the variety and number of SNPs in both our control group and diabetic patients, results were compared by DNAMAN program (**Figure 3**).

According to our results, we observed ten SNPs from nucleotide of 23 of exon 17 between all sequences in this study.

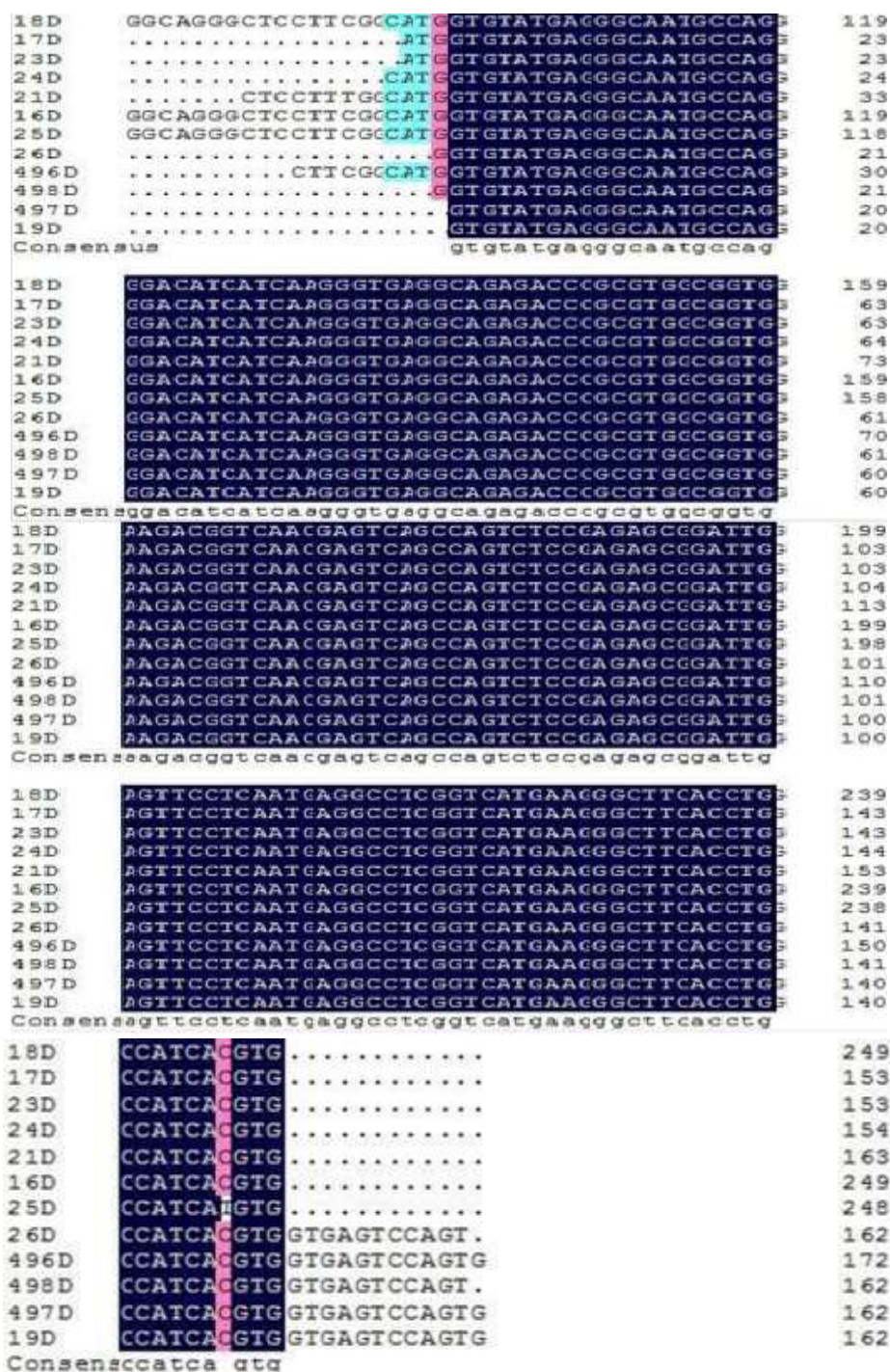


Figure 2: Results of the homology between the sequences of exon 17 in INSR gene in diabetic patients, accession nos. LC055416, LC055417, LC055419, LC055421, LC055423, LC055424, LC055425, LC055426, LC055496, LC055497, and LC055498 by DNAMAN program (D means diabetes and numbers are the end of the accession numbers). According to our results except for the first 21 and the last 12 nucleotides at the end of sequences, other regions of exon 17 has high sequence homology between them.



LC055416ATAATTGGTTCATGCA..CTGTGTAC	GTGCCG	30
LC055417.1CGGTTGTTCATGCT..CTGTGTATA	GTGCCG	31
LC055419 .1	..AATGCAAGTTCCTGCCC..TGTGTAC	GTGCCG	35
LC055421.1	..ATAAGTTCCTGCCC..CTGTGTAC	GTGCCG	34
LC055423.1	..GTAACCAATCTGTCATGCT..CTGTGTAC	GTGCCG	35
LC055424.1	..ATTGTTACGCTTCCACTGCCC..TGTGTAC	GTGCCG	36
LC055425.1ATAATTGGTTCATGCT..CTGTGTAC	GTGCCG	31
LC055426.1ATAGTTCGTTTCATGCT..CTGTGTAC	GTGCCG	32
LC055496.1GGGATCTGTTTCATGCT..CTGTGTAC	GTGCCG	31
LC055497.1ATAGTTCGTTTCATGCT..CTGTGTAC	GTGCCG	33
LC055498.1CGAGGTTTCATGCT..CTGTGTAC	GTGCCG	30
LC055418.1TAGGACAGACATCTGCTCTGTGTAC	GTGCCG	34
LC055420.1	TTAGCGTAACTTCTGCCC..CTGTGTAC	GTGCCG	36
LC055422.1	..ATTGCAAGTTCCTGCCC..TGTGTAC	GTGCCG	34
Consensus		gtgccg	
LC055416	GACGAGTGGGAGGTGTCTCGAGAGAAGATCACCCCTCCTT		69
LC055417.1	GACGAGTGGGAGGTGTCTCGAGAGAAGATCACCCCTCCTT		70
LC055419 .1	GACGAGTGGGAGGTGTCTCGAGAGAAGATCACCCCTCCTT		74
LC055421.1	GACGAGTGGGAGGTGTCTCGAGAGAAGATCACCCCTCCTT		74
LC055423.1	GACGAGTGGGAGGTGTCTCGAGAGAAGATCACCCCTCCTT		74
LC055424.1	GACGAGTGGGAGGTGTCTCGAGAGAAGATCACCCCTCCTT		75
LC055425.1	GACGAGTGGGAGGTGTCTCGAGAGAAGATCACCCCTCCTT		70
LC055426.1	GACGAGTGGGAGGTGTCTCGAGAGAAGATCACCCCTCCTT		71
LC055496.1	GACGAGTGGGAGGTGTCTCGAGAGAAGATCACCCCTCCTT		70
LC055497.1	GACGAGTGGGAGGTGTCTCGAGAGAAGATCACCCCTCCTT		72
LC055498.1	GACGAGTGGGAGGTGTCTCGAGAGAAGATCACCCCTCCTT		69
LC055418.1	GACGAGTGGGAGGTGTCTCGAGAGAAGATCACCCCTCCTT		73
LC055420.1	GACGAGTGGGAGGTGTCTCGAGAGAAGATCACCCCTCCTT		76
LC055422.1	GACGAGTGGGAGGTGTCTCGAGAGAAGATCACCCCTCCTT		74
Consensus	gaagagtgggaggtgtctcgagagaagatcacccctcctt		
LC055416	CGAGAGCTGGGGCAGGGCTCCTTGGCATGGTGTATGAGG		109
LC055417.1	CGAGAGCTGGGGCAGGGCTCCTTGGCATGGTGTATGAGG		110
LC055419 .1	CGAGAGCTGGGGCAGGGCTCCTTGGCATGGTGTATGAGG		114
LC055421.1	CGAGAGCTGGGGCAGGGCTCCTTGGCATGGTGTATGAGG		114
LC055423.1	CGAGAGCTGGGGCAGGGCTCCTTGGCATGGTGTATGAGG		114
LC055424.1	CGAGAGCTGGGGCAGGGCTCCTTGGCATGGTGTATGAGG		115
LC055425.1	CGAGAGCTGGGGCAGGGCTCCTTGGCATGGTGTATGAGG		110
LC055426.1	CGAGAGCTGGGGCAGGGCTCCTTGGCATGGTGTATGAGG		111
LC055496.1	CGAGAGCTGGGGCAGGGCTCCTTGGCATGGTGTATGAGG		110
LC055497.1	CGAGAGCTGGGGCAGGGCTCCTTGGCATGGTGTATGAGG		112
LC055498.1	CGAGAGCTGGGGCAGGGCTCCTTGGCATGGTGTATGAGG		109
LC055418.1	CGAGAGCTGGGGCAGGGCTCCTTGGCATGGTGTATGAGG		113
LC055420.1	CGAGAGCTGGGGCAGGGCTCCTTGGCATGGTGTATGAGG		116
LC055422.1	CGAGAGCTGGGGCAGGGCTCCTTGGCATGGTGTATGAGG		114
Consensus	cgagagctggggcagggctccttggcatgggtgatgagg		
LC055416	GCAATGCCAGGGACATCATCAAGGGTGAGGCAGAGACCCG		149
LC055417.1	GCAATGCCAGGGACATCATCAAGGGTGAGGCAGAGACCCG		150
LC055419 .1	GCAATGCCAGGGACATCATCAAGGGTGAGGCAGAGACCCG		154
LC055421.1	GCAATGCCAGGGACATCATCAAGGGTGAGGCAGAGACCCG		154
LC055423.1	GCAATGCCAGGGACATCATCAAGGGTGAGGCAGAGACCCG		154
LC055424.1	GCAATGCCAGGGACATCATCAAGGGTGAGGCAGAGACCCG		155
LC055425.1	GCAATGCCAGGGACATCATCAAGGGTGAGGCAGAGACCCG		150
LC055426.1	GCAATGCCAGGGACATCATCAAGGGTGAGGCAGAGACCCG		151
LC055496.1	GCAATGCCAGGGACATCATCAAGGGTGAGGCAGAGACCCG		150
LC055497.1	GCAATGCCAGGGACATCATCAAGGGTGAGGCAGAGACCCG		152
LC055498.1	GCAATGCCAGGGACATCATCAAGGGTGAGGCAGAGACCCG		149
LC055418.1	GCAATGCCAGGGACATCATCAAGGGTGAGGCAGAGACCCG		153
LC055420.1	GCAATGCCAGGGACATCATCAAGGGTGAGGCAGAGACCCG		156
LC055422.1	GCAATGCCAGGGACATCATCAAGGGTGAGGCAGAGACCCG		154
Consensus	gcaatgccagggacatcatcaagggtgaggcagagacccg		
LC055416	CGTGGCGGTGAAGACGGTCAACGAGTCAGCCAGTCTCCGA		189
LC055417.1	CGTGGCGGTGAAGACGGTCAACGAGTCAGCCAGTCTCCGA		190
LC055419 .1	CGTGGCGGTGAAGACGGTCAACGAGTCAGCCAGTCTCCGA		194
LC055421.1	CGTGGCGGTGAAGACGGTCAACGAGTCAGCCAGTCTCCGA		194
LC055423.1	CGTGGCGGTGAAGACGGTCAACGAGTCAGCCAGTCTCCGA		194
LC055424.1	CGTGGCGGTGAAGACGGTCAACGAGTCAGCCAGTCTCCGA		195
LC055425.1	CGTGGCGGTGAAGACGGTCAACGAGTCAGCCAGTCTCCGA		190
LC055426.1	CGTGGCGGTGAAGACGGTCAACGAGTCAGCCAGTCTCCGA		191
LC055496.1	CGTGGCGGTGAAGACGGTCAACGAGTCAGCCAGTCTCCGA		190
LC055497.1	CGTGGCGGTGAAGACGGTCAACGAGTCAGCCAGTCTCCGA		192
LC055498.1	CGTGGCGGTGAAGACGGTCAACGAGTCAGCCAGTCTCCGA		189
LC055418.1	CGTGGCGGTGAAGACGGTCAACGAGTCAGCCAGTCTCCGA		193
LC055420.1	CGTGGCGGTGAAGACGGTCAACGAGTCAGCCAGTCTCCGA		196
LC055422.1	CGTGGCGGTGAAGACGGTCAACGAGTCAGCCAGTCTCCGA		194
Consensus	cgaggcggtgaagacgggtcaacgagtcagccagtctccga		
LC055416	GAGCGGATTGAGTTCCTCAATGAGGCCTCGGTCAIGAAGG		229
LC055417.1	GAGCGGATTGAGTTCCTCAATGAGGCCTCGGTCAIGAAGG		230
LC055419 .1	GAGCGGATTGAGTTCCTCAATGAGGCCTCGGTCAIGAAGG		234
LC055421.1	GAGCGGATTGAGTTCCTCAATGAGGCCTCGGTCAIGAAGG		234
LC055423.1	GAGCGGATTGAGTTCCTCAATGAGGCCTCGGTCAIGAAGG		234
LC055424.1	GAGCGGATTGAGTTCCTCAATGAGGCCTCGGTCAIGAAGG		235
LC055425.1	GAGCGGATTGAGTTCCTCAATGAGGCCTCGGTCAIGAAGG		230
LC055426.1	GAGCGGATTGAGTTCCTCAATGAGGCCTCGGTCAIGAAGG		231
LC055496.1	GAGCGGATTGAGTTCCTCAATGAGGCCTCGGTCAIGAAGG		230
LC055497.1	GAGCGGATTGAGTTCCTCAATGAGGCCTCGGTCAIGAAGG		232
LC055498.1	GAGCGGATTGAGTTCCTCAATGAGGCCTCGGTCAIGAAGG		229
LC055418.1	GAGCGGATTGAGTTCCTCAATGAGGCCTCGGTCAIGAAGG		233
LC055420.1	GAGCGGATTGAGTTCCTCAATGAGGCCTCGGTCAIGAAGG		236
LC055422.1	GAGCGGATTGAGTTCCTCAATGAGGCCTCGGTCAIGAAGG		234
Consensus	gagcggtgagttcctcaatgaggcctcgggtcaigaagg		

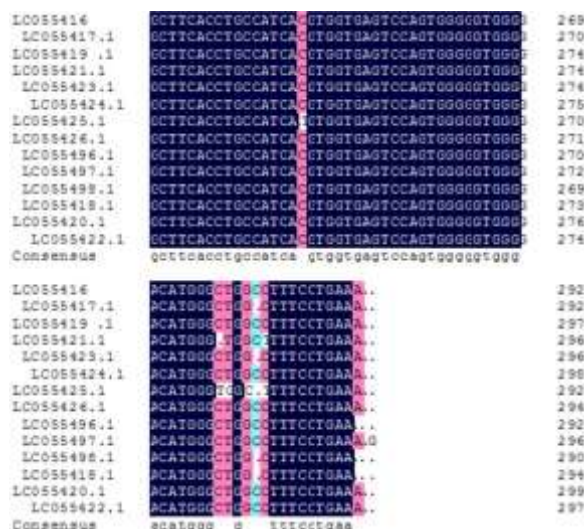


Figure 3: Results of comparison of exon 17 of INSR gene in control and diabetic groups, accession nos. LC055418.1, LC055420 and LC055422.1 (Control's group), LC055416, LC055417, LC055419, LC055421, LC055423, LC055424, LC055425, LC055426, LC055496, LC055497, and LC055498 (diabetic's group) by DNAMAN program. According to our results we observed ten SNPs from nucleotide 23 of exon 17 in all sequences in this study.

In **Figure 4** we also compared a sequence of diabetic patients, accession no. LC055421, with INSR gene accession no. NG_008852.1.

Results only showed one SNP in position 16 of exon 17 in INSR gene.

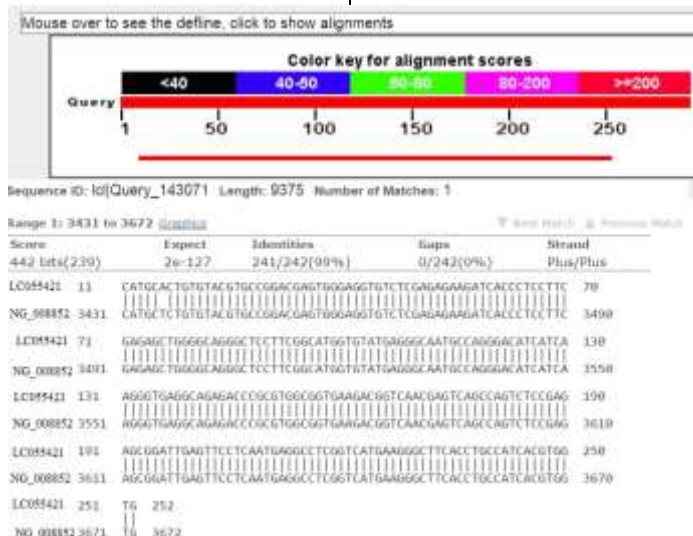


Figure 4. Comparison of sequences with accession no. LC055421 and NG_008852. Results showed that there were a SNP in position 16 of exon 17 in INSR gene.

In this study we investigated single-nucleotide polymorphisms (SNPs) in exon 17 of insulin receptor (INSR) gene in patients with insulin resistance compared to a control group in an Iranian population. Thirty Iranian female patients with diabetics and thirty healthy Iranian women as controls were recruited.

All sixty samples were sequenced but just fourteen samples randomly selected for depositing in Genbank.

Our results showed a significant homology between diabetics and control group in exon 17 of INSR gene when compared by DNAMAN program.

SNPs analysis of exon 17 of INSR gene

This study describes the detection of single nucleotide polymorphisms in exon 17 of INSR gene between diabetic and control groups. Jin and collab. [JIN et al., 2006] have reported a novel T-to-C



substitution at codon Cys1008 (position 3128 in the nucleotide sequence of NM_000208 of INSR). The novel SNP is within exon 17 of INSR, exactly at ATP binding site of the tyrosine kinase domain of INSR β -subunit, resulting in a missense mutation from cysteine to arginine. In current research we studied alignment of exon 17 in INSR gene accession numbers LC055420.1 and NM_000208 to find SNPs between both sequences. According to our results, we found SNPs in position of 3436 for NM_000208 and position of 16 for INSR gene accession no. LC055421.1 (Figure 4). This is an interesting result since Siegel and collab. found a SNP in position of 3128 for exon 17 of INSR gene that is different from our results. On other hand, studies have shown that absence of silent C/T SNP at codon His 1058 has a significant association with PCOS [SIEGEL *et al.*, 2002; WOOD *et al.*, 2003]. To understand the situation of variation between cases and control group, first we separated the subjects into two groups (healthy and diabetics groups). Figure 1 presents healthy group, and as we can see, except for the first 18 nucleotides, three SNPs were observed between the subjects of this group. Here we wanted to know the amount of variation between healthy group subjects collected from different regions of Iran. In Figure 2, exon 17 of INSR in diabetics group were compared by DNAMAN program; the results showed that except for the first 20 nucleotides and the last 12 nucleotides, there is only one SNP in this exon. Variations exist between diabetic group subject may be dependent of geographical regions in Iran. Finally, to understanding the number of nucleotide mutations, we compared both groups (cases and control group).

According to Figure 3, we observed SNPs in different regions of exon 17 in INSR gene. These results showed that in position 55 related to accession no. LC055420.1, LC055421.1 and LC055422.1, there is a single mutation in the form of addition of a C nucleotide, while in other sequences that were studied this nucleotide was absent.

Also we found SNPs in other position of exon 17 in INSR gene that were different between control and diabetic groups. This was a very interesting result because in no other research, like Siegel and Jin [SIEGEL *et al.*, 2002, JIN *et al.*, 2006], such mutations in the same position have been reported. Our results were confirmed NCBI Network system (Figure 4). According to Figure 4, we observed a single mutation in position 11 of exon 17 INSR gene (accession no. LC055421 and NG_008852). Mutations of INSR gene have been reported by other authors [ATIOMO *et al.* 2009; YAMAMOTO *et al.*, 1990, LEGRO *et al.*, 1999, 2004]. They reported that mutation in the insulin receptor can cause a disease with a dominant pattern of inheritance as well. On the other hand, insulin resistance was associated with non-insulin-dependent diabetes mellitus (NIDDM) [MARCOVECCHIO *et al.*, 2005] and may be a central feature of a group of atherogenic metabolic variables sometimes referred to as syndrome X [CHITTENDEN *et al.*, 2009; FARQUHAR *et al.*, 2009; AZIZ, 2002; ODAWARA *et al.*, 1989].

The role of inherited defects of the insulin receptor in these conditions is unknown. Simultaneously, mutations in INSR have been linked to diabetic diseases. Moreover, we found out that the position of single nucleotide mutation between our Iranian population and other populations, such as Chinese population [JIN *et al.*, 2006; BUTU *et al.*, 2015], is different, which may be related to regional variables.

Conclusions

In this research we studied the sequences of INSR gene from Iranian populations. Results showed a variation between control group and diabetic populations. Particularly variations were observed at the beginning and the end of exons. Finally in our next projects we aim to study other exons of INSR for getting a complete picture of the relationship between INSR gene and diabetic diseases.

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Islamic Azad University Tonekabon
Branch, Iran

References

1. Atiomo, W.; Khalid, S.; Parameshweran, S.; Houda, M.; Layfield, R. Proteomic biomarkers for the diagnosis and risk stratification of polycystic ovary syndrome: a systematic review. *BJOG*, **2009**, 116, p. 137–143.
2. Aziz, R. Polycystic ovary syndrome, insulin resistance, and molecular defects of insulin signaling. *Journal of Clinical Endocrinology and Metabolism*. **2002**, 87, p.4085–4087.
3. Butnariu, M.; Sarac, I.; Pentea, M.; Samfira, I.; Negrea, A.; Motoc, M.; Buzatu, A.R.; Ciopec M. Approach for Analyse Stability of Lutein from *Tropaeolum majus*, *Revista de chimie*, **2016**, 67(3), p. 503–506.
4. Butu, A.; Rodino, S.; Golea, D.; Butu, M.; Negoescu, C.; Dinu–Pirvu, C.E.; Butnariu, M. Liposomal nanodelivery system for proteasome inhibitor anticancer drug bortezomib. *Farmacia*, **2015**, 63(2), p.224–229.
5. Chittenden, B.G.; Fullerton, G.; Maheshwari, A.; Bhattacharya, S. Polycystic ovary syndrome and the risk of gynecological cancer: a systematic review. *Reproductive Biomedicine Online*, **2009**, 19(3), p.398–405.
6. Diao, F.Y.; Xu, M.; Hu, Y.; Li, J.; Xu, Z.; Lin, M.; Wang, L.; Zhou, Y.; Zhou, Z.; Liu, J. The molecular characteristics of polycystic ovary syndrome (PCOS) ovary defined by human ovary cDNA microarray. *Journal of Molecular Endocrinology*. **2004**, 33, p.59–72.
7. Dunaif, A. Polycystic ovary syndrome in 2011. Genes, aging and sleep apnea in polycystic ovary syndrome. *Nature reviews*. **2012**, 8(2): 72–74.
8. Eckmann, K.R.; Kockler, D.R. Aromatase inhibitors for ovulation and Farquhar, C.; Lilford, R.J.; Marjoribanks, J.; Vandekerckhove, P. Laparoscopic 'drilling' by diathermy or laser for ovulation induction in anovulatory polycystic ovary syndrome. *Cochrane Database of Systematic Reviews*, **2007**, 18(3), p.11–22.
9. Hoffman, P.L.; Caulfield, W.S.; Robinson, E.M.; Bergman, R.N.; Menon, R.K.; Spelling, M.A.; Gluckman, P.D. Insulin resistance in short children with intrauterine growth retardation. *Journal of Clinical Endocrinology and Metabolism*. **1997**, 82, p. 402–406.
10. Kadowaki, T.; Kadowak, H.; Matthew, M.; Richter, T.; Roth, J.; Gordon, P. et al. Five Mutant Alleles of the Insulin Receptor Gene in Patients with Genetic Forms of Insulin Resistance. *Journal of Clinical Investigation*, **1990**, 86, p. 254–264.
11. Knochenhauer, E.S.; Key, T.J.; Kahsar–Miller, M.; Waggoner, W.; Boots, L.R.; Azziz, R. Prevalence of the polycystic ovary syndrome in unselected black and white women of the southeastern United States: a prospective study. *Journal of Clinical Endocrinology and Metabolism*, **1998**, 83, p. 3078–3082.
12. Lee, E.J.; Yao, K.J.; Kim, S.J.; Lee, S.H.; Chan, K.Y.; Baek, K.H. Single nucleotide polymorphism in exon 17 of the insulin receptor gene is not associated with polycystic ovary syndrome in a Korean population, *Fertile. Sterile*, **2006**, 86, p. 380–384.
13. Legro, R.S.; Castracane, V.D.; Kauffman, R.P. Detecting insulin resistance in polycystic ovary syndrome: purposes and pitfalls, *Obstetrical & gynecological survey*, **2004**, 59, p. 141–154.
14. Legro, R.S.; Kunesman, A.R.; Dodson, Y.W.C.; Dunaif, A. Prevalence and predictors of risk for type 2 diabetes mellitus and impaired glucose tolerance in polycystic ovary syndrome: a prospective, controlled study in 254 affected women. *Journal of Clinical Endocrinology and Metabolism*. **1999**, 84, p.165–169.
15. Li, J.; Xiao, M.Z.; Qiong, L.; Yuli, Q.; Fan, J.; He–Feng, H. A novel SNP at exon 17 of INSR is associated with decreased insulin sensitivity in Chinese women with PCOS. *Molecular Human Reproduction*, **2006**, 12, p.151–155.
16. Li, Y.; Feng, H.L.; Cao, Y.J.; Zheng, G.J.; Yang, Y.; Mullen, S. et al. Confocal microscopic analysis of the spindle and chromosome configurations of human oocytes matured in vitro, *Fertile. Sterile*, **2006**, 85, p. 827–832.
17. Longo, N.; Singh, R.; Griffin, L.D.; Langley, S.D.; Parks, J.S.; Elsas, L.J. Impaired growth in Rabson–Mendenhall syndrome: lack of effect of growth hormone and insulin–like growth factor–I, *Journal of Clinical*



- Endocrinology and Metabolism*, **1994**, 79, p.799–805.
18. Marcovecchio, M.; Mohn, A.; Chiarelli, F. Type 2 diabetes mellitus in children and adolescents. *Journal of Endocrinological Investigation*. **2005**, 28(9), p.853–863.
19. Moeller, D.E.; Yokota, A.; Ginsburg–Fellner, F.; Flier, J.S. Functional properties of a naturally occurring Trp1200 to Ser1200 mutation of the insulin receptor. *Molecular Endocrinology*, **1990**, 4, p. 1183–91.
20. Odawara, M.; Kadowaki, T.; Yamamoto, R.; Shibasaki, Y.; Tobe, K.; Accili, D.; Bevins, C.; Mikami, Y.; Matsuura, N.; Akanuma, Y. Human diabetes associated with a mutation in the tyrosine kinase domain of the insulin receptor. *Science*, **1989**, 245: 66–68.
21. Sambrook, J.; Fritsch, E.F.; Maniatis, T. *Molecular Cloning: a laboratory manual*. 2nd ed. N.Y., Cold Spring Harbor Laboratory, Cold Spring Harbor Laboratory Press, **1998**; p. 1659.
22. San Milan, J.L.; Croton, M.; Villuendas, G.; Sancho, J.; Pearl, J.; Escobar–Morrell, H.F. Association of the polycystic ovary syndrome with genomic variants related to insulin resistance, type 2 diabetes mellitus, and obesity. *Journal of Clinical Endocrinology and Metabolism*, **2004**, 89, p. 2640–2646.
23. Siegel, S.; Futterweit, W.; Davies, T.F.; Conception, E.S.; Greenberg, D.A.; Villanueva, R.; Tomer, Y. A C/T single nucleotide polymorphism at the tyrosine kinase domain of the insulin receptor gene is associated with polycystic ovary syndrome. *Fertile Sterile*, **2002**, 78, p. 1240–1243.
24. Sino, S.; Sino, M.; Bell, G. Human insulin–receptor gene: partial sequence and amplification of exons by polymerase chain reaction. *Diabetes*, **1990**, 39, p. 123–28.
25. Talbot, J.; Bicknell, E.; Rajkhowa, M.; Krook, A.; O'Rahilly, S.; Clayton, R. Molecular scanning of the insulin receptor gene in women with polycystic ovarian syndrome. *Journal of Clinical Endocrinology and Metabolism*, **1996**, 81, p.1979–1983.
26. Tucci, S.; Futterweit, W.; Conception, E.S.; Greenberg, D.A.; Villanueva, R.; Davies, T.F.; Tomer, Y. Evidence for association of polycystic ovary syndrome in caucasian women with a marker at the insulin receptor gene locus. *Journal of Clinical Endocrinology and Metabolism*. **2001**, 86, p. 446–449.
27. Ward, C.W.; Lawrence, M.C.; Streltsov, V.A.; Adams, T.E.; McKeon, N.M. The insulin and EGF receptor structures: new insights into ligand–induced receptor activation. *Trends in Biochemical Science*. **2007**, p.129–137.
28. Weedon, M.N.; Schwarz, P.E.; Horikawa, Y.; Iwasaki, N.; Illig, T et al. Meta–analysis and a large association study confirm a role for calpain–10 variation in type 2 diabetes susceptibility. *American Journal of Human Genetics*, **2003**, 73, p. 1208–1212.
29. Wood, J.R.; Nelson, V.L.; H, C.; Jansen, E.; Wang, C.Y.; Urbane, M.; McAllister, J.M.; Mosselman, S.; Strauss, J.F. The molecular phenotype of polycystic ovary syndrome (PCOS) theca cells and new candidate PCOS genes defined by microarray analysis. *Journal of Biological Chemistry*. **2003**, 278 (29), p. 26380–26390.
30. Xita, N.; Tsatsoulis, A.; Chatzikyriakidou, A.; Giorgio, I. Association of the (TAAAA)_n repeat polymorphism in the sex hormone–binding globulin(SHBG) gene with polycystic ovary syndrome and relation to SHBG serum levels. *Journal of Clinical Endocrinology and Metabolism*, **2003**, 88, p. 5976–5980.
31. Yamamoto, R.; Shiba, T.; Tobe, K.; Shibasaki, Y.; Koshio, O.; Izumi, T.; Odawara, M.; Y. Mikami.; Matsuura, N.; Akanuma, Y. Defect in tyrosine kinase activity of the insulin receptor from a patient with insulin resistance and acanthosis nigricans. *Journal of Clinical Endocrinology and Metabolism*, **1990**, 70, p. 869–878.

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