



CHEMICAL CHARACTERIZATION OF POLYPHENOLS EXTRACTED FROM DIFFERENT HONEYS

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Abstract. The chemical composition of honey varies according to the quality of the harvested nectar. The aim of the study was to determine the qualitative and quantitative composition of honey from different regions in Algeria. A qualitative and quantitative analysis was carried out on honey polyphenol extract with the characterization of the majors groups and the determination of the amount of total polyphenols. The results indicate that the honey polyphenol extracts have different rates of phenolic compounds (Flavonoids, Tannins and Cardenolides). The polyphenol extracts present a concentration of 0.32, 0.4, 0.76 and 0.39 mg GAE/100 g for the extract of Mascara, Oran, Jijel and Tlemcen respectively. The different types of tested honey present a variable composition on phenolic compounds, which allowed us to determine the different therapeutic properties of honey.

Keyword: Honey, Polyphenols, Flavonoids, Tannins, Cardenolides.

Introduction

Nowadays, honey is one of the last remaining untreated natural foods, minimally affected by industrial technologies and highly varying consumer characteristics.

Honey is one of the oldest and traditional sweetening agents for foods and it has retained a "natural" image [APARNA and RAJALAKSHMI, 1999].

Honey is a sweet and flavorful natural product, which has been consumed for its high nutritive value and its contribution in human health [GHELDOLF et al., 2002].

The composition of honey is rather variable and depends primarily on its floral source; however, certain external factors, such as seasonal and environmental factors and processing also play a role.

Honey is a supersaturated solution of sugars, of which fructose (38%) and glucose (31 %) are the main contributors, with phenolic compounds, minerals, proteins, free amino acids, enzymes, and vitamins acting as minor components [ALVAREZ-SUAREZ et al., 2010].

Among minor honey constituents, polyphenols such as flavonoids and phenolic acids may function as natural

antioxidants in our diet [JIALAL and GRUNDY, 1993].

Honey contains phenolic compounds, which are recognised as natural antioxidants thus supporting the application of its use in specific health conditions. These compounds can also be used as the indicators in the studies of the floral and geographical origin of honey and propolis.

Phenolic acids are important group of compounds with respect to appearance and functional properties.

Phenolic compounds occurring in honey have been classified into three groups: flavonoids, cinnamic acids and benzoic acids [AMIOT et al., 1989, [BUTNARIU, et al., 2011, BUTU, et al., 2014b, CAUNII, et al., 2015, BUTNARIU, et al., 2015a].

The aim of the study was to determine the qualitative and quantitative composition of honey from different regions in Algeria.

Material and methods

Extraction of phenolic compounds: 15 g of honey was mixed with hydrochloric acid (6M).

Then, it hydrolyzed in 100 mL of ethanol-water [70–30; V/V]. The solution was added to 2.5 mL of Sodium metabisulfite.



It was followed by stirring for 5 min and filtration. After drying at rotary evaporator at 40 to 70°C, the viscous residue was obtained with present the phenolic extract [SEKOU *et al.*, 2009].

Characterization of phenolic compounds

Flavonoids: The method used was called (Shibata reaction). 2 mL of 10 % polyphenol extract was added to 5 mL of hydrochloric alcohol and 2 or 3 chips of magnesium. An orange or purplish pink color appearing that there was flavonoids [RIBEREAU, 1968, BUTNARIU, *et al.*, 2006, PETRACHE, *et al.*, 2014, BUTNARIU, *et al.*, 2005, BUTU, *et al.*, 2015, BUTNARIU, 2012, RODINO, *et al.*, 2014, BUTNARIU, *et al.*, 2016].

Tannins: The reaction was expressed by the action of ferric chloride (FeCl₃) 5% on the extract of honey 10%. The appearance of a dark blue color or green indicated the presence of tannins [GHAZI and SAHRAOUI, 2005].

Cardenolides: 1 mL of the extract of honey was mixed with 10 mL (CHCl₃ and ethanol). The organic phase was evaporated; the remainder was dissolved in 3 mL of acetic acid.

A few drops of FeCl₃ and 1 mL of concentrated sulfuric acid were added. The colored solution by a blue green color indicated the presence of Cardenolides.

Total Phenolic Content

Principle: Folin Ciocalteu was a yellow acid consisting of a mixture of phosphotungstic acid (H₃PW₁₂O₄₀) and phosphomolybdic acid (H₃PMo₁₂O₄₀). It was reduced in the oxidation of phenols in a mixture of the blue oxides of tungsten and molybdenum [RIBEREAU, 1968, BUTNARIU and

GIUCHICI, 2011, BUTU, *et al.*, 2014a, BUTNARIU, 2014, IANCULOV, *et al.*, 2004, BUTNARIU, *et al.*, 2015b].

The color produced in $\lambda = 760$ nm was proportional to the amount of polyphenols present in plant extracts [GHAZI and SAHRAOUI, 2005].

Protocol: The measuring of Total Phenolic Content was carried out by using a SHIMADZU UV-2401PC UV spectrophotometer with $\lambda = 760$ nm.

For the determination of total polyphenols, we used the Folin-Ciocalteu. 0.2 mL of the honey polyphenol extract was added to 1 mL of Folin Ciocalteu reagent with Bicarbonate (CO₃Na₂) 4.25 %.

The control solution was prepared with distilled water. Then, the solutions were brought in a water bath at 70°C for 20 minutes.

After cooling, the optical density was determined at 760 nm compared to the control.

The use of a standard range was established under the same conditions with Gallic acid (0 to 1 g / L).

The Folin Ciocalteu was expressed in gram of Gallic acid / L.

To ensure that the results were reliable, the dosage of each phenolic compound was carried out in three trials.

Then, the average optical density was calculated [GHAZI and SAHRAOUI, 2005].

Results and discussion

Polyphenolic extract: Each sample of honey has recovered a precise amount of polyphenolic extract (table 1).

Table 1.

Color and aspect of honey extracts.

Samples	Colors	Aspect	Yields (%)
Jijel	Brown	Viscous	18.5 ± 2
Oran	Brown	Viscous	20 ± 2.9
Tlemcen	Yellow	Liquid	13 ± 1.5
Mascara	Yellow	Liquid	20.2 ± 2.3

Characterization of Phenolic Compounds:

Characterization of phenolics compounds of honeys allowed detecting qualitatively the types of polyphenols (table 2).

From the table below, we see the abundance of flavonoids in the four samples especially in honey of Jijel and Oran explained by the brown color of these extracts.



Flavonoids vary quantitatively and qualitatively according to season and stage of plant development [FRONTY, 2008].

However, tannins and cardenolides were scarce.

This amount was related to the botanical origin of each variety of honey [WHITE, 1980, [BUTNARIU and CORADINI, 2012, BARBAT, et al., 2013, BUTNARIU, et al., 2012, BUTU, et al., 2014c].

Table 2.

Phenolic compounds detected in honey extracts

Compounds	Identification	Jijel	Oran	Mascara	Tlemcen
Flavonoids	Orange / purplish pink	+++	++	+	++
Tanins	Dark blue / green	+	+	-	-
Cardenolides	Blue green	-	+	+	-

Dosage of polyphenols: After preparing the calibration range of Gallic acid (0.25 g / L; 0.5 g / L; 0.75 g / L; 1g / L), the measurement of the optical density was performed at $\lambda=760$ nm.

The absorbance obtained was plotted against concentrations; the

calibration curve obtained showed the linearity of the detector response as a function of different concentrations.

The mean absorbance of polyphenolic extracts of honey (Jijel, Mascara, Oran, Tlemcen) were shown in table 3.

Table 3.

The optical densities of polyphenolic extracts.

Extract	Jijel	Oran	Mascara	Tlemcen
Optical Density ($\lambda=760$ nm)	0.721	0.401	0.286	0.360

The concentration of each extract was 0.32, 0.4, 0.76 and 0.39 mg GAE/100 g for the extract of Mascara, Oran, Jijel and Tlemcen respectively (figure 1).

This large difference was probably due to the storage conditions of the original honey and by the different solids content of honey.

According to Harris [HARRIS, 1977], these different levels of phenolic

compounds in the different extracts of honey were the result of the effect of influential factors on the honeys, the main ones: light, precipitation, the rate of HMF and season.

In this regard, [MACHEIX et al., 1990] reported that the concentration of polyphenols varied from one variety to another and declined steadily during ripening and storage.

Table 4.

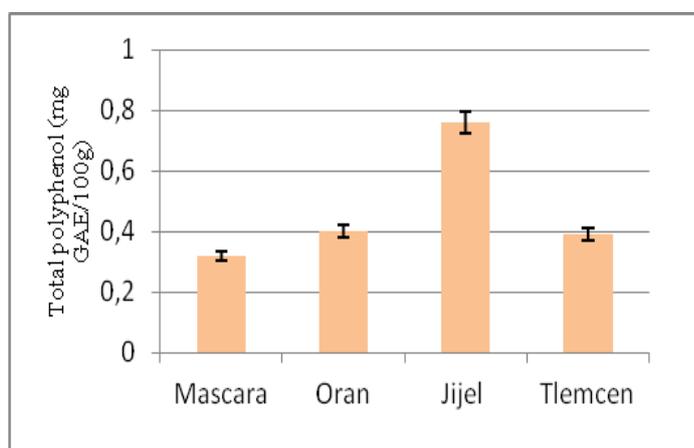


Figure 1. Rate of total polyphenols of different honeys.

This variation was based on several factors including temperature, pH and the quantitative and qualitative changes in

phenolic compounds contained in the original plants [AMIOT et al., 1995].



Conclusions

The results of this study indicate that the honey polyphenol extracts have different rates of phenolic compounds (Flavonoids, Tannins and Cardenolides).

It presents a concentration of 0.32, 0.4, 0.76 and 0.39 mg GAE/100 g for the extract of Mascara, Oran, Jijel and Tlemcen respectively.

The different types of honey present a variable composition on phenolic compounds which allowed us to determine the different therapeutic properties of honey.

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References

1. Alvarez-Suarez, J.M.; Tulipani, S.; Romandini, S.; Bertoli, E.; Battino, M. Contribution of honey in nutrition and human health: a review. *Mediterranean Journal of Nutrition and Metabolism*, **2010**, 3, 15–23.
2. Amiot, M.J.; Aubert, S.; Gonnet, M.; and Tacchini, M. Phenolic compound of honeys: Preliminary study on the identification and quantification by families. *Apidologie*, **1989**, 20, 115–125.
3. Amiot, M.J.; Tacchini, M.; Aubert, S.Y.; Olezek, W. Influence of cultivar, maturity stage, and storage condition on phenolic composition and enzymatic browning of pear fruits. *Journal of Agricultural and Food Chemistry*, **1995**, 43 (5), 1132–1137.
4. Aparna, A.R.; Rajalakshmi, D. Honey—Its characteristics, sensory aspects and applications. *Food Reviews International*, **1999**, 15, 455–471.
5. Barbat, C.; Rodino, S.; Petrache, P.; Butu, M.; Butnariu, M. Microencapsulation of the allelochemical compounds and study of their release from different, *Digest journal of nanomaterials and biostructures*, **2013**, 8(3), 945–953.
6. Butnariu, M. An analysis of *Sorghum halepense*'s behavior in presence of tropane alkaloids from *Datura stramonium* extracts, *Chemistry central journal*, **2012**, 6(75).
7. Butnariu, M. Detection of the polyphenolic components in *Ribes nigrum* L. *Annals of agricultural and environmental medicine*, **2014**, 21(1), 11–4.
8. Butnariu, M.; Bostan, C. Antimicrobial and anti-inflammatory activities of the volatile oil compounds from *Tropaeolum majus* L. (Nasturtium), *African journal of biotechnology*, **2011**, 10(31), 5900–5909.
9. Butnariu, M.; Caunii, A.; Putnoky, S. Reverse phase chromatographic behaviour of major components in *Capsicum Annuum* extract, *Chemistry central journal*, **2012**, 6(146).
10. Butnariu, M.; Coradini, C.Z., Evaluation of Biologically Active Compounds from *Calendula officinalis* Flowers using Spectrophotometry, *Chemistry central journal*, **2012**, 6(35).
11. Butnariu, M.; Goian, M.; Ianculov, I.; Gergen, I.; Negrea, P. Studies about CO²⁺ ion influence on soy plants development and acumulation of other chemical elements (Iron, magnesium, calcium, potassium and phosphorus), *Revista de chimie*, **2005**, 56(8), 837–841.
12. Butnariu, M.; Negrea, P.; Lupa, L.; Ciopec, M.; Negrea, A.; Pentea, M.; Sarac, I.; Samfira, I. Remediation of Rare Earth Element Pollutants by Sorption Process Using Organic Natural Sorbents. *International journal of environmental research and public health*, **2015**, 12(9), 11278–11287b.
13. Butnariu, M.; Samfira, I.; Sarac, I.; Negrea, A.; Negrea, P. Allelopathic effects of *Pteridium aquilinum* alcoholic extract on seed germination and seedling growth of *Poa pratensis*, *Allelopathy journal*, **2015**, 35(2), 227–236a.
14. Butnariu, M.; Sarac, I.; Pentea, M.; Samfira, I.; Negrea, A.; Motoc, M.; Buzatu, A.R.; Ciopec, M. Approach for Analyse Stability of Lutein from *Tropaeolum majus*, *Revista de chimie*, **2016**, 67(3), 503–506.
15. Butnariu, M.; Smuleac, A.; Dehelean, C.; Chirita, R.; Saratean, V. Studies concerning fertilizer influence (NPK in different doses) on quantity of corn plants chlorophyll, *Revista de chimie*, **2006**, 57(11), 1138–1143.
16. Butnariu, M.V.; Giuchici, C.V. The use of some nanoemulsions based on aqueous propolis and lycopene extract in the skin's protective mechanisms against UVA radiation,



- Journal of nanobiotechnology*, **2011**, 9(3).
17. Butu, A.; Rodino, S.; Golea, D.; Butu, M.; Butnariu, M.; Negoescu, C.; Dinu-Pirvu, C.E. Liposomal nanodelivery system for proteasome inhibitor anticancer drug bortezomib, *Farmacia*. **2015**, 63(2), 224–229.
18. Butu, M.; Butnariu, M.; Rodino, S.; Butu, A. Study of zingiberene from *Lycopersicon esculentum* fruit by mass spectrometry, *Digest journal of nanomaterials and biostructures*, **2014**, 9(3), 935–941c.
19. Butu, M.; Rodino, S.; Butu, A.; Butnariu, M. Screening of bioflavonoid and antioxidant activity of *Lens culinaris medikus*, *Digest journal of nanomaterials and biostructures*, **2014**, 9(2), 519–529a.
20. Butu, M.; Rodino, S.; Pentea, M.; Negrea, A.; Petrache, P.; Butnariu, M. IR spectroscopy of the flour from bones of European hare, *Digest journal of nanomaterials and biostructures*. **2014**, 9(4), 1317–1322b.
21. Caunii, A.; Butu, M.; Rodino, S.; Motoc, M.; Negrea, A.; Samfira, I.; Butnariu, M. Isolation and Separation of Inulin from *Phalaris arundinacea* Roots, *Revista de chimie*, **2015**, 66(4), 472–476.
22. Fronty, F. Honey and its benefits. *Paris, flammation*, **2008**, 86.
23. Ghazi, F.; Sahraoui, S. Evolution of phenolic compounds and total carotenoids during maturation of two varieties of dates Tantboucht and Hamraia, thesis of agronomy, El Harrach, **2005**.
24. Gheldof, N.; Xiao-Hong, W.; Engeseth, N. Identification and quantification of antioxidant components of honeys from various floral sources, *Journal of Agricultural and Food Chemistry*, **2002**. 50, 5870–5877.
25. Harris, R. Nutritional evaluation of food processing, The Avi Publishing Company Inc., **1977**, Westport, CT.
26. Ianculov, I.; Gergen, I.; Palicica, R.; Butnariu, M.; Dumbrava, D.; Gabor, L. The determination of total alkaloids from *Atropa belladonna* and *Lupinus sp* using various spectrophotometrical and gravimetrical methods, *Revista de chimie*, **2004**, 55(11), 835–838.
27. Jialal, I.; Grundy, S.M. Effect of supplementation with alpha tocopherol, ascorbate and beta-carotene on low-density lipoprotein oxidation. *Circulation*, **1993**, 88, 2780–2786.
28. Macheix, J.J.; Fleuret, A.; Billot, J. Fruit's phenolics. CRC Press, Inc., Raton, **1990**.
29. Petrache, P.; Rodino, S.; Butu, M.; Pribac, G.; Pentea, M.; Butnariu, M. Polyacetylene and carotenes from *Petroselinum sativum* root, *Digest journal of nanomaterials and biostructures*, **2014**, 9(4), 1523–1527.
30. Ribereau, G.P. Plant phenolic compounds. Dunod, Paris, 254 p, **1968**.
31. Rodino, S.; Butu, M.; Negoescu, C.; Caunii, A.; Cristina, R.T.; Butnariu, M. Spectrophotometric method for quantitative determination of nystatin antifungal agent in pharmaceutical formulations, *Digest journal of nanomaterials and biostructures*, **2014**, 9(3), 1215–1222.
32. Sekou, D.; Kouakou, E.K.; Ouolo, A.; Coulibal, H.F. Performance of two extraction techniques of root phenols for the evaluation of the marking tolerance to Fusarium wilt of Oil palm clones (*Elaeis guineensis* Jacq.). *Sciences & Nature*, **2009**, 6(2), 117–123.
33. White, J. Report on the analysis of honey, ASSOC.OFF Agric, *Chemists* 34–48 p, **1980**.

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