



Investigation of the induced antibiosis resistance by zinc element in different cultivars of sugar beet to long snout weevil, *Lixus incanescens* (Col: Curculionidae)

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Abstract. Sugar beet weevil, *Lixus incanescens* Boh., is one of most dangerous pests in sugar beet. Host plant resistance is known as an efficient strategy for pest management. Thus, this study was conducted to investigate the antibiosis resistance induced by zinc element in different cultivars of sugar beet to long snout weevil, *L. incanescens*. Seeds of eleven commercial monogerm cultivars of sugar beet including Pars, Torbat, Ekbatan, SBSI019, Rizofort, Puma, Dorothea, Brigita, Pecora, Nagano and Jolgeh, were studied. Field was divided to two parts; one part was treated with zinc fertilizer and other part did not treat with fertilizer. Antibiosis parameters were tested. Results clearly did not show that cultivars have antibiosis resistance under non-using fertilizer during 2013 and 2014 years. Rizofort cultivar showed lower susceptibility compared with other cultivars when using zinc fertilizer, because of lower adult weight, female longevity and period length of immature stage during the both years. Thus, Rizofort cultivar was relatively resistant to *L. incanescens* when using zinc fertilizer during the both years, which can be useful in the development of IPM programs for this insect in sugar beet fields

Keyword: *Lixus incanescens*, antibiosis, sugar beet, zinc fertilizer.

Introduction

Sugar beet is belonging to *Beta vulgaris* L. ssp. *vulgaris* sugar beet group [LANGE *et al.*, 1999]. Sugar beet is extensively cultivated in northern hemisphere in late winter or early spring and harvested after 5–9 months, depending on climatic and soil conditions. Twenty percent of world's supply of sugar is achieved from sugar beet and other part is get from sugar cane which is mainly cultivated in tropical climates in developing countries [FAO, 2009].

The sugar content in sugar beet has different amounts. The by-products of the sugar beet including pulp and molasses increases value of up to 10 % of the value of the sugar. However, sugar beet was protected by herbicides [WINNER, 1993].

Sugar beet is severely sensitive to pests when developing the first 3–4 pairs of leaves [BAZOK *et al.*, 2016]. Sugar beet weevil, *Lixus incanescens* Boh., is one of most dangerous pests in sugar beet which is found in some parts of Iran and

other countries. This pest attacks leaf and petiole of sugar beet and larvae can damage up to 75 % root weight loss [ARBABTAFTI *et al.*, 2012; OCETE *et al.*, 1994]. It is well known that the use of insecticides for controlling the insects has caused some problems such as the development of insecticide resistance, health hazards, and the risk of environmental pollution [COX, 2004]. Farmers usually use pesticides for controlling this pest, but often these efforts are not successful, since immature stages are located the petioles [FOSTER *et al.*, 2007], resulting increased resistance of *L. incanescens*. However, the use of insecticides is still efficient strategies for controlling the pest [ROUHANI *et al.*, 2012].

Discovery of some genetic resistances to diseases enhanced sugar yield and lowered reliance on pesticides. Studies have shown that improvement in hybrid varieties, the pest and disease resistances, breeding assisted by molecular biology, etc. caused to produce



hybrid varieties. Host plant resistance is known as an efficient strategy for pest management, because of its importance economic and environment [KARIMI *et al.*, 2012, SAMFIRA *et al.*, 2015].

Plant species are known to have different suitability for different insect pests when their performance and preference are compared with other plants [RAZMJOU and GOLIZADEH, 2010; RAZMJOU *et al.*, 2012]. Antixenosis, antibiosis, and tolerance are resistance mechanisms known against insects in plants [KOGAN and ORTMAN, 1978; MOTTAGHINIA *et al.*, 2011]. Studies have shown that variations in host plant quality can influence development, reproduction and, hence, population growth of target herbivorous insects [La ROSSA *et al.*, 2013].

Some active ingredients are allowed to control sugar beet weevil, because of implementation of EU pesticide legislation. Zinc can be used to control the pest, since it is known to have antimicrobial activity which can be applied as fungicide [SEVEN *et al.*, 2004; LEVIN *et al.*, 2007].

Zinc acts as antimicrobial agent by generation of reactive oxygen species, oxidative stress, membrane disruption, protein unfolding, and/or inflammation [REDDY *et al.*, 2007; MENG *et al.*, 2009; DONALDSON *et al.*, 2009]. It is also reported the effects of zinc against insects (*Aphis nerii*) [ROUHANI *et al.*, 2012]. Other study has shown impacts of zinc oxide nanoparticle against rice weevil (Coleoptera: Curculionidae) [KERATUM *et al.*, 2015, CAUNII, *et al.*, 2015, BUTNARIU, *et al.*, 2015].

The use of different varieties of sugar beet with diverse quantitative and qualitative characteristics can increase yield and decrease pest losses in the world. So far, based on our knowledge no comprehensive study in Iran is done about induction of antibiosis resistance by zinc in sugar beet varieties to sugar beet long snout weevil. Thus, this study aimed to investigate the induction of antibiosis resistance by zinc in eleven sugar beet varieties to long snout weevil *L. incanescens* in Iran.

Material and methods

Location and Plant materials. The present study was conducted at Pistachio Research Station, Yezbar village Qazvin

Province, Iran (36° 2'N, 50° 7'E, 1175 m above sea level). Seeds of eleven commercial monogerm cultivars of sugar beet including Pars, Torbat, Ekbatan, SBSI019, Rizofort, Puma, Dorothea, Brigita, Pecora, Nagano and Jolgeh, were prepared from Seed and Plant Breeding Research Institute, Karaj, Iran.

Preparing land and insect colony. Field testing was achieved in an area of about 0.1 hectare, in the early spring of 2013 and 2014 that was deeply plowed. After tight and weeding, planting seeds of sugar beet varieties were conducted in the second decade of April each year.

Farm irrigation every 7 to 10 days were routine. Weed control was performed by hand removing. Adult's weevils were collected from a sugar beet (*B. vulgaris*) field.

Field preparation for planting and zinc sulphate treatment. Field was divided to two parts; one part was treated with zinc fertilizer and other part did not treat with fertilizer. Each plot was divided to 3 blocks and each block was also divided to 3 plots. Each plot had 11 m length and 5.5 m width and 11 plots were in each row. Plot distance was 1 m and row distance were 11 m.

Antibiosis experiment. After 30 days and to develop 3 to 8 leaves, part of plant in main plot was selected and maintained at cages with dimensions 50·30·30 cm. Insects (3 males and 2 females) with <1 d of age was released into each cage. Insects were weighted and allowed to feeding up to 7 days and were then weighted. This action was in 3 replicates.

Longevity and also larval weight was calculated.

Statistical analysis. The data are shown as mean \pm standard deviation (mean \pm SD). Prior to analysis, heterogeneity of variance was tested and then all entries were transformed [Log(X+2)]. Numbers of adult weevils deployed at the different times on eleven sugar beet cultivars at two-years were analyzed based on combined analysis of randomized complete block design.

Significance of the differences was examined by analysis of variance (ANOVA) test., followed using Tukey post



hoc. The levels of significance were done at $\alpha=0.05$. Statistical analysis was performed by SAS[®] 9.1 software (2005).

Results and discussion

Mean comparison of treatments in 2013 year (Table 1) showed that when using zinc fertilizer, the lowest insect weight was observed in Rizofort cultivar and the highest insect weight was observed in Brigita cultivar ($P<0.05$).

However, when non-using zinc fertilizer, the highest insect weight was observed in Pars and Puma and the lowest were considered in Nagano cultivar ($P<0.05$) in 2013 year (Table 2).

Considering males longevity, Jolgeh cultivar had highest male longevity and puma showed the lowest male longevity (Table 1), while when non-using zinc fertilizer, Pars showed highest male longevity (Table 2).

Table 1.

Means comparison of antibiotics test in different cultivars when using zinc fertilizer in 2013 year (Means \pm SE)

Cultivars	Adult weight (mg)	Male longevity (d)	Female longevity (d)	Period length of immature stages (d)	Larval weight (mg)
Pars	0.011 \pm 0.001bcd	6.66 \pm 0.33cd	11.00 \pm 0.57a	28.00 \pm 0.57a	0.015 \pm 0.000a
Torbat	0.012 \pm 0.001bcd	7.66 \pm 0.33bc	10.00 \pm 0.57ab	26.66 \pm 0.88a	0.013 \pm 0.000a
Ekbatan	0.009 \pm 0.00cde	8.33 \pm 0.33abc	9.66 \pm 0.33ab	29.66 \pm 0.88a	0.012 \pm 0.001a
SBSI019	0.013 \pm 0.001abc	7.66 \pm 0.33bc	8.66 \pm 0.33ab	27.33 \pm 0.88a	0.012 \pm 0.000a
Rizofort	0.006 \pm 0.000e	7.33 \pm 0.33bcd	7.66 \pm 0.33b	28.33 \pm 0.33a	0.015 \pm 0.000a
Puma	0.008 \pm 0.000de	5.66 \pm 0.33d	9.00 \pm 0.00ab	26.33 \pm 1.20a	0.015 \pm 0.000a
Drothea	0.012 \pm 0.001bcd	8.00 \pm 0.00abc	8.66 \pm 0.33ab	27.00 \pm 1.15a	0.012 \pm 0.001a
Brigita	0.016 \pm 0.000a	7.66 \pm 0.33bc	10.66 \pm 0.33ab	30.33 \pm 1.20a	0.012 \pm 0.000a
Pecora	0.015 \pm 0.000ab	9.00 \pm 0.00ab	10.66 \pm 0.33ab	30.33 \pm 1.20a	0.012 \pm 0.001a
Nagano	0.014 \pm 0.000ab	7.66 \pm 0.33bc	9.00 \pm 0.00ab	29.33 \pm 1.85a	0.015 \pm 0.000a
Jolgeh	0.014 \pm 0.000ab	9.66 \pm 0.33a	10.33 \pm 1.20ab	29.00 \pm 1.15a	0.011 \pm 0.000a

Mean values in a column followed by different lowercase letters (a–e) are significantly different on the basis of ANOVA with Tukey's test ($p<0.05$)

Considering female longevity, when using zinc fertilizer, Pars cultivar had the highest longevity, but it was not observed significant differences when non-using zinc sulphate fertilizer.

There was no significant difference among groups for period length of

immature stage during two years ($P>0.05$).

When using zinc fertilizer, it was not found significant differences among groups for larval weight, however, Rizofort cultivar showed the highest for when non-using zinc fertilizer.

Table 2.

Means comparison of antibiotics test in different cultivars when non-using zinc fertilizer in 2013 year (Means \pm SE)

Cultivars	Adult weight (mg)	Male longevity (d)	Female longevity (d)	Period length of immature stages (d)	Larval weight (mg)
Pars	0.024 \pm 0.001a	8.33 \pm 0.33a	9.00 \pm 0.57a	25.00 \pm 0.57a	0.016 \pm 0.001ab
Torbat	0.015 \pm 0.000bc	7.33 \pm 0.33ab	8.66 \pm 0.33a	26.33 \pm 0.88a	0.016 \pm 0.000ab
Ekbatan	0.020 \pm 0.001ab	6.66 \pm 0.33ab	8.66 \pm 0.88a	26.00 \pm 0.57a	0.018 \pm 0.001ab
SBSI019	0.016 \pm 0.000bc	6.00 \pm 0.00b	8.33 \pm 0.33a	24.66 \pm 0.66a	0.017 \pm 0.000ab
Rizofort	0.016 \pm 0.001bc	6.66 \pm 0.33ab	8.66 \pm 0.33a	25.33 \pm 0.88a	0.021 \pm 0.001a
Puma	0.025 \pm 0.000a	6.33 \pm 0.33b	9.33 \pm 0.33a	25.00 \pm 0.00a	0.015 \pm 0.001b
Drothea	0.018 \pm 0.001bc	6.00 \pm 0.00b	8.00 \pm 0.57a	25.66 \pm 1.15a	0.017 \pm 0.000ab
Brigita	0.016 \pm 0.000bc	7.33 \pm 0.33ab	9.00 \pm 0.57a	24.00 \pm 0.57a	0.015 \pm 0.000ab
Pecora	0.017 \pm 0.001bc	7.00 \pm 0.57ab	7.66 \pm 0.33a	25.00 \pm 0.57a	0.018 \pm 0.000ab
Nagano	0.014 \pm 0.001c	6.33 \pm 0.33b	9.00 \pm 0.00a	26.33 \pm 0.66a	0.015 \pm 0.000b
Jolgeh	0.016 \pm 0.000bc	6.33 \pm 0.33b	7.66 \pm 0.33a	27.00 \pm 1.00a	0.016 \pm 0.001ab

Mean values in a column followed by different lowercase letters (a–c) are significantly different on the basis of ANOVA with Tukey's test ($p<0.05$)

The mean for antibiotics tests for different cultivars in 2014 year are shown in Table 3. When application of zinc fertilizer, Pecora and Jolgeh showed the

highest insect weight and Puma showed lowest, however, Nagano showed highest insect weight when non-application the insect.



Table 3.

Means comparison of antibiosis test in different cultivars when using zinc fertilizer in 2014 year (Means ± SE)

Cultivars	Adult weight (mg)	Male longevity (d)	Female longevity (d)	Period length of immature stages (d)	Larval weight (mg)
Pars	0.015±0.001bcd	7.33±0.33bcd	10.00±0.00abc	29.00±0.57cd	0.012±0.001a
Torbat	0.016±0.000ab	8.00±0.00abc	11.00±0.57ab	30.33±0.88bcd	0.014±0.000a
Ekbatan	0.013±0.00bc	8.66±0.33abc	10.33±0.66abc	31.66±0.88abc	0.012±0.000a
SBSI019	0.012±0.000c	7.33±0.33bcd	8.33±0.33cde	26.66±0.88de	0.013±0.000a
Rizofort	0.012±0.000c	7.00±0.57cd	7.00±0.00e	24.00±0.57e	0.016±0.000a
Puma	0.008±0.000d	5.66±0.33d	9.00±0.00bcde	29.33±0.88bcd	0.014±0.000a
Drothea	0.012±0.000c	8.66±0.33abc	7.66±0.33de	32.33±0.33abc	0.012±0.000a
Brigita	0.013±0.000bc	9.00±0.00ab	9.66±0.33abcd	27.66±0.33de	0.013±0.001a
Pecora	0.017±0.000a	7.66±0.33abc	10.66±0.33ab	33.00±0.57ab	0.014±0.001a
Nagano	0.016±0.000ab	7.33±0.33bcd	11.33±0.66a	35.33±0.881a	0.012±0.001a
Jolgeh	0.017±0.000a	9.33±0.33a	9.66±0.33abcd	27.66±0.88de	0.011±0.000a

Mean values in a column followed by different lowercase letters (a–e) are significantly different on the basis of ANOVA with Tukey's test (p<0.05)

With regards to males longevity, Puma cultivar showed the lowest male longevity (Table 3), while when non–using zinc fertilizer, while Drothea showed lowest male longevity (Table 4). Considering female longevity, when using

zinc fertilizer, Rizofort cultivar had the lowest longevity, while when non–using zinc sulphate fertilizer, Pecora cultivar showed the lowest significant differences. It was not found significant differences among groups for larval weight (P>0.05).

Table 4.

Means comparison of antibiosis test in different cultivars when non–using zinc fertilizer in 2014 year (Means ± SE)

Cultivars	Adult weight (mg)	Male longevity (d)	Female longevity (d)	Period length of immature stages (d)	Larval weight (mg)
Pars	0.027±0.001ab	7.00±0.57ab	8.66±0.33a	24.66±0.88a	0.016±0.000a
Torbat	0.019±0.004bc	6.66±0.66ab	7.66±0.33ab	26.33±0.33a	0.015±0.000a
Ekbatan	0.021±0.000bc	7.00±0.57ab	8.66±0.33a	25.00±0.57a	0.018±0.001a
SBSI019	0.014±0.000c	6.33±0.66ab	8.00±0.00ab	25.33±1.45a	0.017±0.001a
Rizofort	0.024±0.002bc	6.00±0.00ab	8.33±0.66ab	26.00±0.57a	0.021±0.000a
Puma	0.021±0.001bc	5.66±0.33ab	9.00±0.57a	25.66±0.33a	0.015±0.002a
Drothea	0.019±0.000bc	5.33±0.66b	9.00±0.57a	25.33±0.88a	0.016±0.001a
Brigita	0.023±0.001bc	6.00±0.00ab	9.00±0.00a	24.66±88a	0.015±0.000a
Pecora	0.019±0.001bc	7.66±0.33a	6.00±0.57b	25.33±0.33a	0.017±0.001a
Nagano	0.035±0.003a	6.66±0.33ab	7.66±0.66ab	25.66±0.88a	0.017±0.000a
Jolgeh	0.017±0.002bc	6.33±0.33ab	7.66±0.33ab	28.33±0.33a	0.019±0.000a

Mean values in a column followed by different lowercase letters (a–c) are significantly different on the basis of ANOVA with Tukey's test (p<0.05)

The results of data analysis for the mechanism of antibiosis test in the year 2013 year (Table 5) and 2014 year (Table

6) show significant differences between experimental for cultivars tested.

Table 5.

Combined analysis of antibiosis test when using zinc fertilizer in different sugar beet cultivars during 2013 and 2014 years

S.O.V.	Df	Adult weight (mg)	Male longevity (d)	Female longevity (d)	Period length of immature stages (d)	Larval weight (mg)
Year	1	0.000064**	0.060 NS	0.060 NS	29.333**	0.000000 ns
Block (year)	4	0.000000 NS	0.424 NS	0.530 NS	2.424 NS	0000000 NS
Cultivar	10	0.000044**	6.003**	7.469**	21.442**	0.00001**
Year × Cultivar	10	0.000010**	0.760*	1.627 NS	16.133**	0.000003 NS
Error	40	0.000001	0.340	0.880	2.624	0.0000028
C.V.	–	10.348	7.497	9.829	5.574	12.720

NS: non–significant, *: p<0.05 and **: p<0.01



There was significant interaction between year and cultivar for insect weight ($P < 0.05$).

The sugar beet is known to be one of the most important productions in farming fields in Qazvin province, Iran.

This study investigated antibiosis responses of eleven sugar beet cultivars against *L. incanescens*. The results of present study showed a wide range of host plant responses to sugar beet long snout weevil between sugar beet cultivars when using and non-using zinc fertilizer.

The results of this study can help to use the best cultivar in selection of suitable cultivar, regarding to longevity and period length of immature stages.

Antibiosis test can be used as efficient in preventing their populations from reaching economic damage levels [KENNEDY *et al.*, 1987, IANCULOV, *et al.*, 2004, BUTU, *et al.*, 2014]. To evaluate the resistance by population growth parameters of an insect is beneficial tools in showing degree of plant resistance to insects [TSAI and WANG, 2001, RASHED and BUTNARIU, 2014]. In 2013 year, Rizofort cultivar showed lower insect weight, female longevity and period length of immature stage. However, it did not show significant difference with those which had male longevity for male longevity traits when using zinc fertilizer. It can be stated that Rizofort cultivar has lower susceptibility compared with other cultivars when using zinc fertilizer.

Table 6.

Combined analysis of antibiosis test when non-using zinc fertilizer in different sugar beet cultivars during 2013 and 2014 years

S.O.V.	Df	Adult weight (mg)	Male longevity (d)	Female longevity (d)	Period length of immature stages (d)	Larval weight (mg)
Year	1	0.000232 **	1.833 NS	2.560 NS	0.545 NS	0.0000000 NS
Block (year)	4	0.000021 NS	0.757 NS	0.242 NS	0.803 NS	0000002 NS
Cultivar	10	0.000068 **	2.078 **	2.581 **	4.678 *	0.000016 **
Year × Cultivar	10	0.000067 **	0.733 NS	0.793 NS	0.745 NS	0.000001 NS
Error	40	0.000008	0.474	0.692	1.703	0.0000040
C.V.	–	14.494	10.448	9.967	5.102	11.762

NS: non-significant, *: $p < 0.05$ and **: $p < 0.01$

At same year, Brigita had the highest insect weight and period length of immature stage. It seems that Brigita cultivar has higher sensitivity when using zinc fertilizer. Insects reared on Rizofort had lowest female longevity and period length of immature stages [STOLERU *et al.*, 2018].

Thus, it decreases reproduction. However, Nagano cultivar showed lower insect weight and male longevity and Pars cultivar showed the highest insect weight and male longevity when non-using.

Thus, to treat with zinc can influence resistance in different cultivars. The decreased insect weight can implicit on antibiosis mechanism [HORTON *et al.*, 1997].

In 2014 Fathi and Abedi showed that Period length of immature stages were different among sugar beet cultivars [FATHI and ABEDI, 2014]. In the study published by Golizadeh and colleagues on five sugar beet cultivars (Doroti, Perimer, Pershia, Rozier and 006) found relative high levels of susceptibility in Pershia

cultivar to *Aphis fabae*; however, the cultivars Rozier and Perimer were relatively less susceptible [GOLIZADEH *et al.*, 2016, PETRACHE, *et al.*, 2014, BUTNARIU, *et al.*, 2016].

On the basis our findings, under application and non-application of zinc fertilizer, the different cultivars showed the different antibiosis resistance. It seems that different cultivars have various interactions with zinc. It is shown that heavy metals such zinc in insects have a clear effect on growth [WARRINGTON, 1987], mortality [MITTERBOCK and FUHRER, 1988], and physiology [ILIJIN *et al.*, 2009, BUTU, *et al.*, 2014, RODINO, *et al.*, 2014, BUTNARIU and SAMFIRA, 2012].

Thus, zinc can help to antibiosis resistance in different cultivars. At 2014 year, Rizofort cultivar again showed lower susceptibility compared with other cultivars when using zinc fertilizer, because it had lower insect weight, female longevity and period length of immature stage.



During 2013 and 2014 years, this cultivar showed better efficiency for antibiosis test. Thus, Rizofort cultivar and the use of zinc can be suggested as resistance cultivar.

It was studied the effects of four cultivars of sugar beet on biology and demographic parameters of *A. fabae* and showed that the use of Shirin cultivar could upgrade the natural biological control of *A. fabae* by *Aphidius matricariae* [TAHRIRI ADABI *et al.*, 2010].

Our findings also indicated that the antibiosis parameters of *L. incanescens* were differentially affected by the sugar when non-using zinc fertilizer.

The data about the quality of sugar beet cultivars and their interactions with fertilizers can affect the antibiosis parameters for *L. incanescens* and it also can help us understand the population dynamics and may help in the development of better management programs for this pest.

However, Rizofort cultivar can be beneficial when using zinc fertilizer. Our observations showed that the Rizofort cultivar was relatively resistant to *L. incanescens* when using zinc fertilizer, which can be useful in the development of IPM programs for this insect in sugar beet fields.

Conclusions

This study was conducted to investigate the antibiosis resistance induced by zinc element in different cultivars of sugar beet to long snout weevil. Our findings clearly did not show that cultivars have antibiosis resistance under non-using fertilizer.

However, Rizofort cultivar was relatively resistant to *L. incanescens* when using zinc fertilizer, which can be useful in the development of IPM programs for this insect in sugar beet fields.

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References

1. Arbabtafti, R.; Sheikhi Garjan, A.; Hosseini Gharalari, A. Crop loss assessment of *Lixus incanescens* Boh. (Coleoptera: Curculionidae) on sugar beet, *Beta Vulgaris* L. *Jordan Journal of Biological Sciences*, **2012**; 3: 197–202.
2. Bazok, R.; Šatvar, M.; Radoš, I.; Drmi, Č.Z.; Lemić, D.; Čačija, M.; Viri, Č.; Gašparić, H. Comparative Efficacy of Classical and Biorational Insecticides on Sugar Beet Weevil, *Bothynoderes punctiventris* Germar (Coleoptera: Curculionidae). *Plant Protection Science*, **2016**; 52 (2): 134–141.
3. Butnariu, M.; Samfira, I. Free Radicals and Oxidative Stress. *Journal of Bioequivalence and Bioavailability*, **2012**, 4, 0975–0851
4. Butnariu, M.; Samfira, I.; Sarac, I.; Negrea, A.; Negrea, P. Allelopathic effects of *Pteridium aquilinum* alcoholic extract on seed germination and seedling growth of *Poa pratensis*. *Allelopathy journal*, **2015**, 35(2), 227–236a.
5. Butnariu, M.; Sarac, I.; Pentea, M.; Samfira, I.; Negrea, A.; Motoc, M.; Buzatu, A.R.; Ciopec, M. Approach for Analyse Stability of Lutein from *Tropaeolum majus*. *Revista de chimie*, **2016**, 67(3), 503–506.
6. Butu, M.; Butnariu, M.; Rodino, S.; Butu, A. Study of zingiberene from *Lycopersicon esculentum* fruit by mass spectrometry. *Digest journal of nanomaterials and biostructures*, **2014**, 9(3), 935–941c.
7. Caunii, A.; Butu, M.; Rodino, S.; Motoc, M.; Negrea, A.; Samfira, I.; Butnariu, M. Isolation and Separation of Inulin from *Phalaris arundinacea* Roots. *Revista de chimie*, **2015**, 66(4), 472–476.
8. Cox, P.D. Potential for using semichemicals to protect stored products from insect infestation. *Journal of Stored Products Research*, **2004**; 40: 1–25.
9. Donaldson, K.P.J.A.; Borm, V.; Castranova, M.; Gulumian. The limits of testing particle-mediated oxidative stress in vitro in predicting diverse pathologies; relevance for testing of nanoparticles. *Particle and Fibre Toxicology*, **2009**; 6: 1–13.
10. FAO, 2009. Sugar Beet White Sugar, FAO Investment Centre Division, Rome,



- Italy, [Online], Available: [http://www.fao.org/fileadmin/user_upload/tci/docs/AH1\(eng\)_Sugar_%20beet%20white%20sugar.pdf](http://www.fao.org/fileadmin/user_upload/tci/docs/AH1(eng)_Sugar_%20beet%20white%20sugar.pdf) [Accessed: 25.7.2013].
11. Fathi, S.A.A.; Abedi, A.A. Ovipositional preference and life history parameters of *Lixus incanescens* (Coleoptera: Curculionidae) on selected sugar beet cultivars. *International Journal of Pest Management*, **2014**; 60 (4): 293–299.
12. Foster, S.P.; Devine, G.; Devonshire, A.L. Insecticide resistance. In: van Emden, H.F.; Harrington, R. (eds) Aphids as crop pests. *CAB International, Wallingford*, **2007**; 261–278.
13. Golizadeh, A.; Abedi, Z.; Borzoui, E.; Golikhajeh, N.; Jafary, M. Susceptibility of Five Sugar Beet Cultivars to the Black Bean Aphid, *Aphis fabae* Scopoli (Hemiptera: Aphididae). *Neotropical Entomology*, **2016**; 45: 427–432.
14. Horton, D.N.; Chauvin, R.L.; Hinojosa, T.; Larson, D.; Murphy, C; Biever, K.D. Mechanism of resistance to Colorado potato beetle in several potato lines and correlation with defoliation. *Entomologia Experimentalis et Applicata*, **1997**; 82: 239–248.
15. Ianculov, I.; Gergen, I.; Palicica, R.; Butnariu, M.; Dumbrava, D.; Gabor, L. The determination of total alkaloids from *Atropa belladonna* and *Lupinus sp* using various spectrophotometrical and gravimetrical methods. *Revista de chimie*, **2004**, 55(11), 835–838.
16. Ilijin L.; Periać-Mataruga, V.; Radojičić, R.; Lazarević, J.; Nenadović, V.; Vlahović, M.; Mrdaković, M. Effects of cadmium on protocerebral neurosecretory neurons and fitness components in *Lymantria dispar* L. *Folia Biologica, (Krakow)*, **2009**; 58 (1–2): 91–99.
17. Karimi, S.; Fathipour, Y.; Talebi, A.A.; Naseri, B. Evaluation of canola cultivars for resistance to *Helicoverpa armigera* (Lepidoptera: Noctuidae) using demographic parameters. *Journal of Economic Entomology*, **2012**; 105: 2172–2179.
18. Kennedy, G.G.F.; Gould, O.M.; Deponti, B.; Stinner, R.E. Ecological, agricultural, genetic, and commercial considerations in the deployment of insect-resistant germplasm. *Environmental Entomology*, **1987**; 16: 327–338.
19. Keratum, A.Y.; Arab, R.B.; Ismail, A.A.; Nasr, G.M. Impact of nanoparticle zinc oxide and aluminum oxide against rice weevil *Sitophilus oryzae* (coleoptera: curculionidae) under laboratory conditions. *Egy. J. Plant Pro. Res.*, **2015**; 3(3): 30–38.
20. Kogan, M.; Ortman, E.E. Antixenosis: a new term proposed to replace painters non-performance modality of resistance. *Bulletin of the Entomological Society of America*, **1978**; 24:175–176.
21. La Rossa, F.R.; Vasicek, A.; López, M.C. Effects of pepper (*Capsicum annuum*) cultivars on the biology and life table parameters of *Myzus persicae* (Sulz.) (Hemiptera: Aphididae). *Neotropical Entomology*, **2013**; 42: 634–641.
22. Lange, W.; Brandenburg, W.A.; De Bock, T.S.M. Taxonomy and cultonomy of beet (*Beta vulgaris* L.). *Botanical Journal of the Linnean Society*, **1999**; 130: 81–96.
23. Levin, M.D.; Den Hollander, J.G.; B. Van Der Holt, B.; Rijnders, B.J.; Van Vliet, M.; Sonneveld, P.; Van Schaik, R.H. Hepatotoxicity of oral and intravenous voriconazole in relation to cytochrome P450 polymorphisms. *Journal of Antimicrobial Chemotherapy*, **2007**; 60(5): 1104–1107.
24. Meng, H.; Xia, T.; George, S.; Nel, A.E. A predictive toxicological paradigm for the safety assessment of nanomaterials. *ACS Nano*, **2009**; 3: 1620–1627.
25. Mitterbock, F.; Fuhrer, E. Effects of fluoride-polluted spruce leaves on nun moth caterpillars (*Lymantria monacha*). *Journal of Applied Entomology*, **1988**; 105(1): 19–27.
26. Mottaghinia, L.; Razmjou, J.; Nouri-Ganbalani, G.; Rafiee-Dastjerdi, H. Antibiosis and antixenosis of six commonly produced potato cultivars to the green peach aphid, *Myzus Persicae* Sulzer (Hemiptera: Aphididae). *Neotropical Entomology*, **2011**; 40: 380–386.
27. Ocete, R.; Ocete, M.E.; Perez-Izquierdo, M.A.; Rubio, I.M. Approximation to the phenology of *Lixus junci* Boh. (Col.: Curculionidae) in La Rioja Alta: estimate of the damage it causes. *Boletín de Sanidad Vegetal Plagas*, **1994**; 20: 611–616.
28. Petrache, P.; Rodino, S.; Butu, M.; Pribac, G.; Pentea, M.; Butnariu, M.



- Polyacetylene and carotenes from *Petroselinum sativum* root, *Digest journal of nanomaterials and biostructures*, **2014**, 9(4), 1523–1527.
29. Rashed, K.N.; Butnariu, M. Isolation and antimicrobial and antioxidant evaluation of bio-active compounds from *Eriobotrya japonica* stems. *Advanced Pharmaceutical Bulletin*, **2014**, 4(1):75–81.
30. Razmjou, J.; Golizadeh, A. Performance of corn leaf aphid, *Rhopalosiphum maidis* (Fitch) (Homoptera: Aphididae) on selected maize hybrids under laboratory conditions. *Applied Entomology and Zoology*, **2010**; 45: 267–274.
31. Razmjou, J.; Mohamadi, P.; Golizadeh, A.; Hasanpour, M.; Naseri, B. Resistance of wheat Lines to *Rhopalosiphum padi* (Hemiptera: Aphididae) under laboratory conditions. *Journal of Economic Entomology*, **2012**; 105: 592–5.
32. Reddy, K.M.; Feris, K.; Bell, J.; Wingett, D.G.; Hanley, C.; Punnoose, A. Selective toxicity of zinc oxide nanoparticles to prokaryotic and eukaryotic system. *Applied Physics Letters*, **2007**; 90 :2139021–2139023.
33. Rodino, S.; Butu, M.; Negoescu, C.; Caunii, A.; Cristina, R.T.; Butnariu, M. Spectrophotometric method for quantitative determination of nystatin antifungal agent in pharmaceutical formulations, *Digest journal of nanomaterials and biostructures*, **2014**, 9(3), 1215–1222.
34. Rouhani, M.; Samih, M.A.; Kalantari, S. Insecticide effect of silver and zinc nanoparticles against *Aphis nerii* boyer de fonscolombe (hemiptera: aphididae). *Chilean Journal of Agricultural Research*, **2012**: 72(4): 590–594.
35. Samfira, I.; Rodino. S; Petrache. P; Cristina. R.T; Butu. M; Butnariu. M. Characterization and identity confirmation of essential oils by mid infrared absorption spectrophotometry. *Digest journal of nanomaterials and biostructures*. 10(2), **2015**, 557–565.
36. SAS Institute. SAS/STAT user's guide, version 9.1. Cary (NC): SAS Institute, **2005**.
37. Seven, O.; Dindar, B.; Aydemir, S.; Metin, D.; Ozinel, M.A.; Icli, S. Solar photocatalytic disinfection of a group of bacteria and fungi aqueous suspensions with TiO₂, ZnO and Sahara Desert dust. *Journal of Photochemistry and Photobiology A: Chemistry*, **2004**; 165(1–3): 103–107.
38. Stoleru, V.; Munteanu, N.; Stan, T.; Ipatioaie, C.; Cojocaru, L.; Butnariu, M. Effects of production system on the content of organic acids in Bio rhubarb (*Rheum rhabarbarum* L.). *Romanian Biotechnological Letters*, **2018**. DOI:10.26327/RBL2017.98.
39. Tahriri Adabi, S.; Talebi, A.A.; Fathipour, Y.; Zamani, A. Life history and demographic parameters of *Aphis fabae* (Hemiptera: Aphididae) and its parasitoid, *Aphidius matricariae* (Hymenoptera: Aphidiidae) on four sugar beet cultivars. *Acta Entomologica Serbica*, **2010**; 15: 61–73.
40. Tsai, J.H.; Wang, J.J. Effects of host plant on biology and life table parameters of *Aphis spiraecola* (Hom. Aphididae). *Environmental Entomology*, **2001**; 30: 44–50.
41. Warrington, S. Relationship between SO₂ dose and growth of the pea aphid, *Acyrtosiphon pisum*, on peas. *Environmental Pollution*, **1987**; 43(2): 155–162.
42. Winner, C. History of the crop. In: Cooke D.A.; Scott, R.K. (eds.) The sugar beet crop. *Chapmann & Hall, London, UK*, **1993**; 1–35.

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